



Alpbach – 14th March 2007



ESF ProteomeBinders Workshop on Affinity Proteomics:

Anticalins and other alternative scaffolds



Protein Scaffolds for the engineering of novel binding sites





The Lipocalin protein family: prominent members



Lipocalin	Abbreviation	No. of residues	Oligomeric state	Glycosylation	No. of SS-bonds	Physiological ligand(s)
α_1 -Acid glycoprotein	AGP	183	monomer	+	2	? (numerous drugs)
Apolipoprotein D	ApoD*	169	mon./HDL-assoc.	+	2	Progesterone/Arachidonate
Apolipoprotein M	ApoM	188	HDL-anchored	+	3	?
Complement component 8 γ chain	C8 γ *	182	heterodimer	-	1	?
<i>Epididymal lipocalin / E-RABP</i>	LCN6	142	monomer	+	0	Retinoic acid ?
Glycodelin	PP14	162	dimer	+	2	?
α_1 -Microglobulin / protein HC	α_1 m	183	fusion \rightarrow mon./het.	+	1	Heme
Odorant-binding protein	OBP	157	monomer/dimer	-	1	Odorant compounds
Prostaglandin D synthase / β -Trace	PGDS*	169	monomer	+	1	Prostaglandin D/H ₂
Retinol-binding protein	RBP*	183	monomer	-	3	Retinol (Vitamin A)
Siderocalin / NGAL	LCN2*	178	monomer/het.	+	1	Fe(III) siderophore
Tear lipocalin	LCN1/Tlc*	158	monomer/dimer	-	1	Retinoids/Rifampin/Lipids...
β -Lactoglobulin	Blg*	162	monomer/dimer	-	2	Fatty acids/Retinol
Aphrodisin	-	151	monomer	+	2	?
Major urinary protein	MUP*	166	dimer	-	1	2-(s-Butyl)thiazoline etc.
Bilin-binding protein	BBP*	173	dimer (monomer)	-	2	Biliverdin IX _b
Lazarillo	LAZ	171	GPI-linked	+	1-2	?
Bacterial lipocalin	Blc*	159	lipoprotein	-	0	?

(human members of the family)

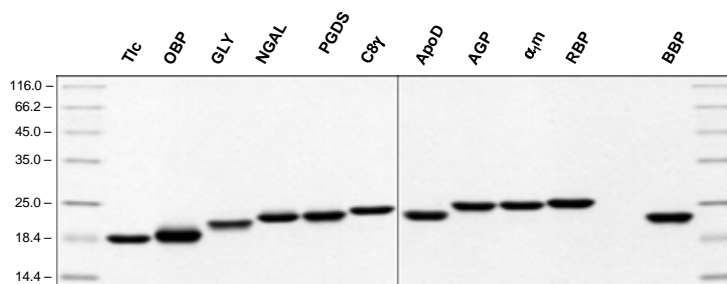
*Crystal structure known



Standardized bacterial production of ten human lipocalins



1. Secretion of recombinant lipocalins into the periplasm of *E. coli* via the OmpA signal peptide
2. Purification from the periplasmic cell fraction via the **Strep-tag II** and gel filtration
3. Coomassie-stained SDS PAGE (15 %):

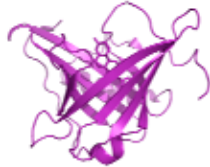




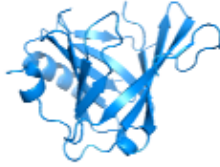
3D gallery of 10 human lipocalins: β -barrel fold with 4 variable loops



AGP



ApoD
[PDB: 2HZQ]



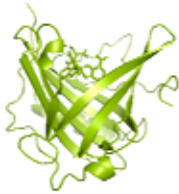
C8 γ
[PDB: 1IW2]



PP14



PGDS
[PDB: 2CZU]



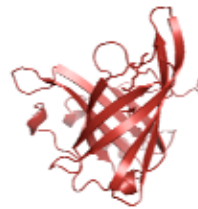
NGAL
[PDB: 1L6M]



OBP



α_m



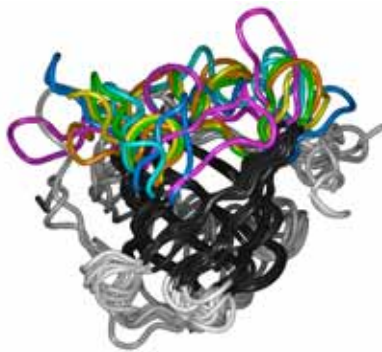
RBP
[PDB: 1RBP]



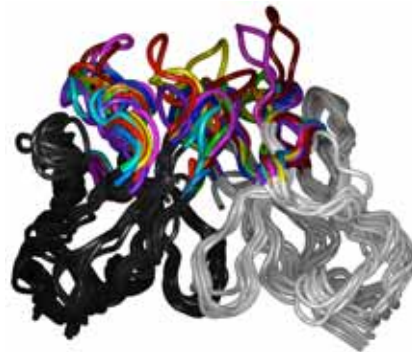
Tlc
[PDB: 1XKI]



Lipocalins & Antibodies: structural variability in the loop region



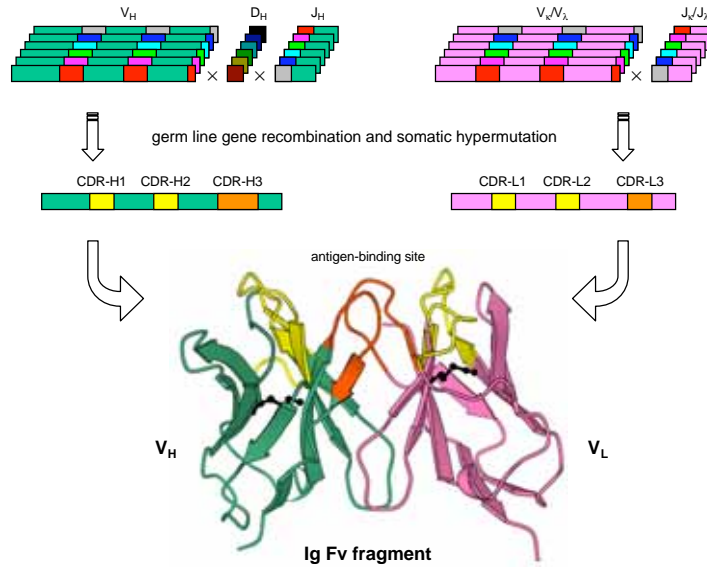
6 "prototypic" lipocalin structures:
RBP BBP MUP EPA BLG BDA



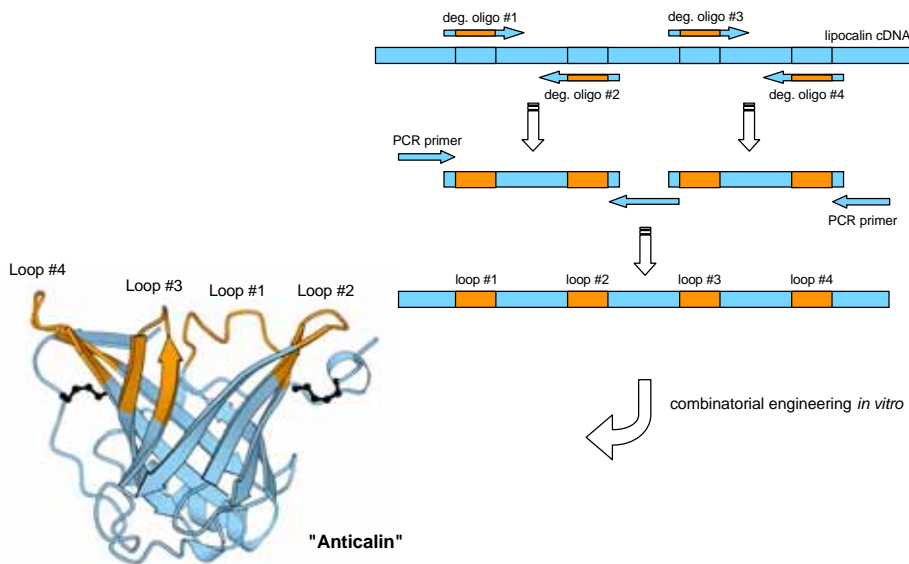
10 Fv fragments of different antibodies



Generation of diversity in the immune system

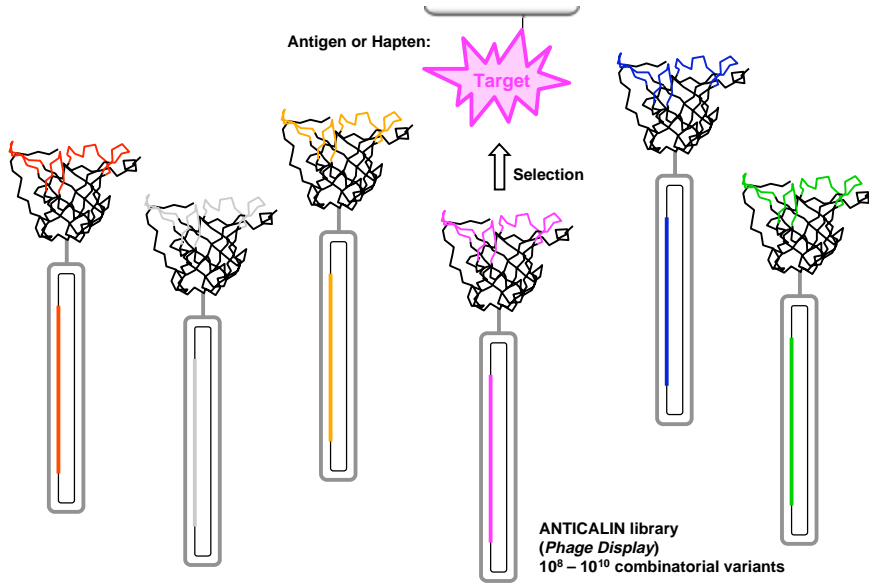


ANTICALIN®s: engineered lipocalins with novel binding properties

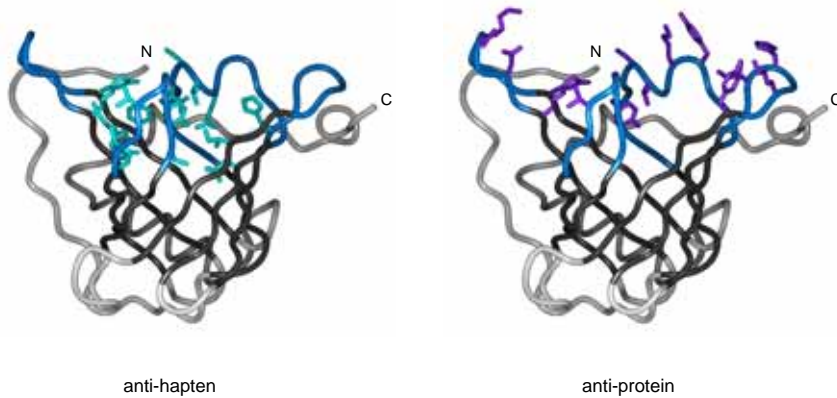




Selection from a lipocalin library with randomized binding sites



BBP random libraries designed against hapten or antigen targets





Targets and applications for ANTICALINS



Established targets for ANTICALINS:

- a) Haptens:
 - fluorescein
 - digoxigenin
 - etc.
- b) Peptides
 - His₆-tag
 - HBV epitope
 - Aβ peptide
- c) Protein antigens (medically relevant), e.g.:
 - CD4
 - CD33
 - **CTLA-4**
 - Cytokine receptor
 - VEGF

Established applications for ANTICALINS:

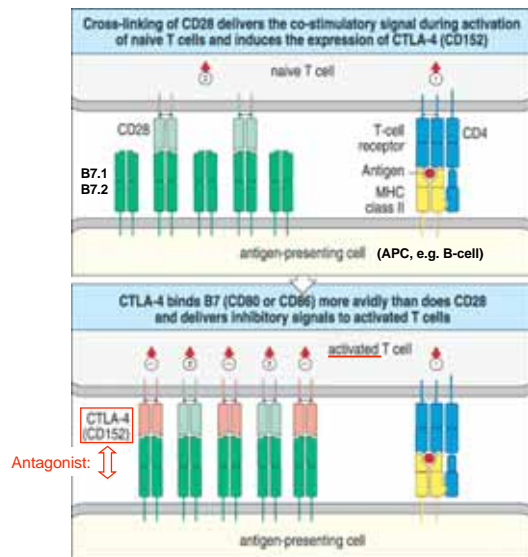
- typical "immunochemical" assays
 - Western blot
 - ELISA
 - BIAcore
 - Flow-cytometric staining of cells
 - Histochemistry
- therapeutic applications ...

ANTICALIN libraries based on human scaffolds:

- **NGAL** (Siderocalin)
- Tear lipocalin
- Apolipoprotein D



Releasing the *immune brake* by an ANTICALIN as CTLA-4 antagonist



- ⇒ Clinical proof of concept for the immunotherapy of solid tumors (metastatic melanoma) with two human(ized) antibodies:
- MDX-010 (Medarex): Phase III
 - CP670 (Pfizer): Phase I/II

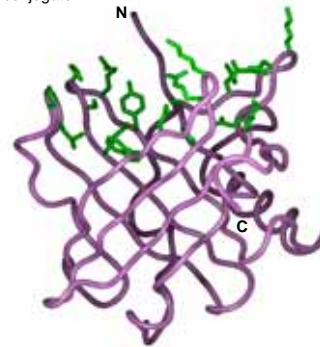
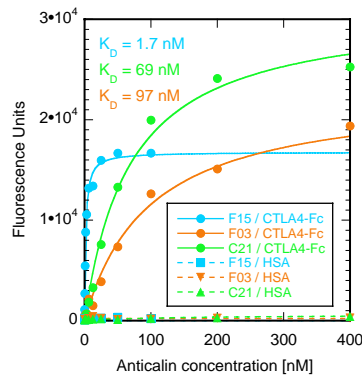
from Janeway et al. "Immunobiology"



Selection of ANTICALINs with high affinity towards CTLA-4



- Selection of from a hNGAL library (20 randomized positions, complexity $2.0 \cdot 10^{10}$) via phage display followed by screening and identification of individual ANTICALINs via ELISA-HTS
- Target: **CTLA4-IgG1** (immunoadhesin of the extracellular T-cell co-receptor **CD152**)
 - a) directly coated, 5 cycles of panning → ANTICALIN "F15"
 - b) capturing via murine α huFc antibody, 5 cycles of panning → ANTICALIN "C21" + "F03"
- ELISA:
 1. capturing (5 μ g/mL murine α huFc antibody) of huCTLA4-Ig
 2. application of soluble, purified ANTICALIN (via *Strep*-tag II and gel filtration)
 3. detection via N-terminal T7-tag with mAb/HRP conjugate



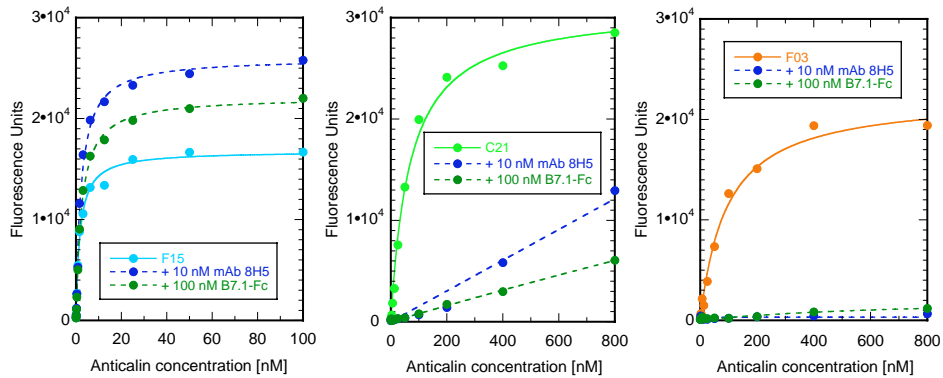
The hNGAL random library



Epitope specificity of α CTLA-4 ANTICALINs



- ELISA with purified ANTICALINs as before, but after pre-incubation with competitors: (detection of bound ANTICALIN via N-terminal T7-tag with mAb/HRP conjugate)

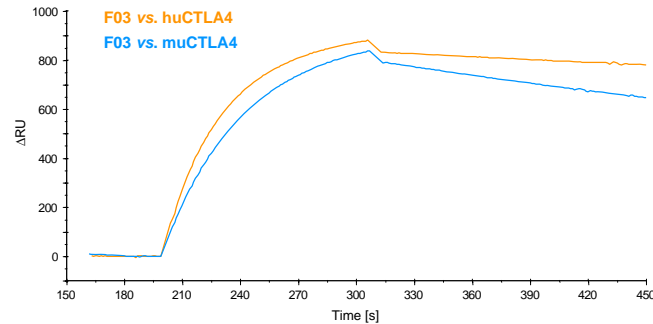




The ANTICALIN "F03" cross-reacts with human and murine CTLA-4



- immobilization of ~2000 RU huCTLA4 or of ~2000 muCTLA4, both as hulgG1 Fc fusion proteins on a CM5 BIACORE chip coated with amine-coupled murine α huFc antibody (14000 RU)
- application of the *Strep*-tag II-purified ANTICALIN F03, here at 5 μ M concentration (20 μ l/min)



Results:

- huCTLA4: $k_{on} = 7.03 \cdot 10^3 \text{ M}^{-1} \text{ s}^{-1}$ $k_{off} = 3.3 \cdot 10^{-4} \text{ s}^{-1}$ $K_D = 47 \text{ nM}$
- muCTLA4: $k_{on} = 4.5 \cdot 10^3 \text{ M}^{-1} \text{ s}^{-1}$ $k_{off} = 8.9 \cdot 10^{-4} \text{ s}^{-1}$ $K_D = 198 \text{ nM}$



Engineering of the α CTLA-4 ANTICALIN for higher affinity & stability



1. Engineering for improved target affinity:

- amplification of the central coding region of the anticalin "F03" via error-prone PCR
- phage display selection against huCTLA4-Ig (3 cycles), followed by filter sandwich colony screen (or ELISA HTS)
- ↳ "J08" (and "Q13")
- ↳ $K_D = 7.9 \text{ nM}$ towards human CTLA-4 / 14.0 nM towards mouse CTLA-4
- ↳ $T_m = 48.3 \text{ }^\circ\text{C}$

2. Engineering for improved thermal stability:

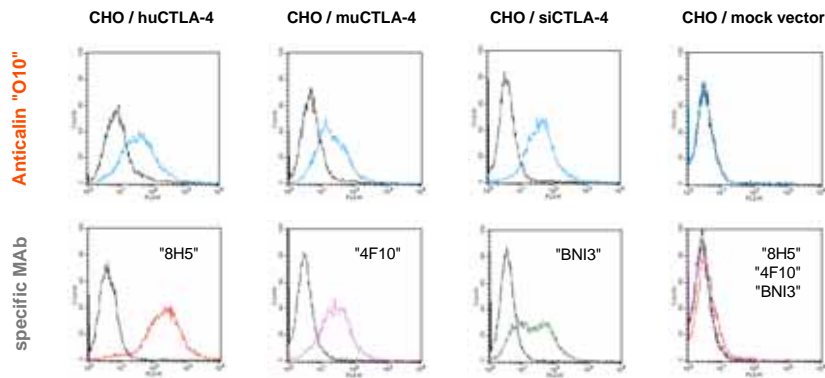
- amplification of the central coding region of the anticalin "J08" via error-prone PCR
- phage display selection against huCTLA4-Ig at increased stringency, including preincubation of phagemids at elevated temperatures ($T = 70 \text{ }^\circ\text{C}$ / 3 cycles), followed by ELISA HTS, in the presence of B7.1-Ig as competitor and/or including preincubation of protein extracts at $60 \text{ }^\circ\text{C}$
- ↳ "O10"
- ↳ $K_D = 3.9 \text{ nM}$ towards human CTLA-4 / 15.3 nM towards murine CTLA-4
- ↳ $T_m = 65.5 \text{ }^\circ\text{C}$



α CTLA-4 ANTICALIN "O10" in flow cytometry (FACS analysis)



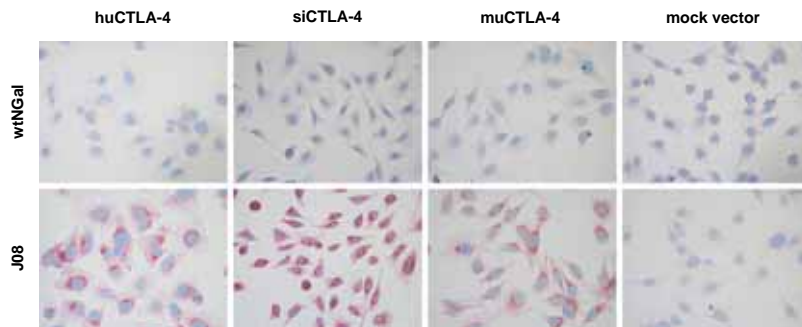
- CHO cells transfected with CTLA-4 cDNA from different species
- incubation of cells with purified ANTICALIN, followed by NGAL-specific mAb "HYB211-02"-Bio and detection via streptavidin-phycoerythrin (PE) conjugate
- alternatively, incubation with various species-specific α CTLA4 MAb (biotinylated or dye-labeled)
(black line: isotype control, i.e. IgG or wtNGAL)



Histochemical staining of CTLA-4 positive cells with "J08"

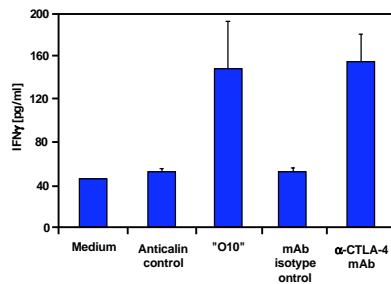


- specific staining of CTLA4-transfected, acetone-fixed adherent CHO cells (cytospin preparation on frozen slides)
- detection of FITC-labelled ANTICALIN via anti-fluorescein mouse antibody and anti-mouse-AP conjugate, followed by FastRed staining and hematoxylin counterstaining





Enhanced T-cell activation *in vitro* by "O10" (PRS010) in MLR assay



Mixed Lymphocyte Reaction (MLR):

against allogeneic (MHC disparate) CD80⁺/CD86⁺ JY cells (human B-cells)

⇒ initial T-cell activation is attenuated by CTLA-4 unless blocked by cognate ANTICALIN or mAb

- co-culture of 10⁵ washed human PHA T-cell blast cells (PBMC + 1 μ g/ml PHA for 72 h) with 5,000 irradiated JY cells for 72 h
- 50 μ g/ml of either CTLA-4 blocking antibodies or ANTICALINS are added at the start of the co-culturing period
- measurement of IFN γ production after 48, 96 h (PHA blasts alone + test substance \approx 20 pg/ml)

enhanced T-cell activity

- as revealed by
- a) interferone production
 - b) cellular proliferation

⇒ potential for treatment of CTCL (Cutaneous T Cell Lymphoma) and immunotherapy of various solid tumors



Value of 'alternative scaffolds' for proteome research?



Functionally:

- selection from synthetic libraries ⇒ control of many intrinsic and extrinsic parameters
- choice of selection techniques: phage display, ribosome display, colony screening, intracellular inhibition etc.
- monoclonality ⇒ reproducibility in binding behaviour and unlimited resources
- controllable specificity, e.g. by subtractive or competitive selection schemes
- binding activity may be enhanced as needed by *in vitro* affinity maturation
- higher robustness, smaller size, easier manufacture
- simple fusion or conjugation with reporter or affinity moieties
- recognition of native vs. denatured targets?
- preference of particular epitope shapes for different scaffolds?



Survey of engineered protein scaffolds for molecular recognition



	Name	Template	Size	No. of loops	Crosslinks	Commercial exploitation
Ig-like domains	Nanobody	cameloid VHH	~ 130	3	2 S-S	www.ablynx.com
	dAb	human single V _H	~ 120	3	1 S-S	www.domantis.com
	iMab	Ig type fold	~ 110	3 – (4+4)	1 S-S	www.catchmabs.com
	Adnectin	¹⁰ F _n 3 fibronectin III	94	2	–	www.adnexustx.com
loopy folds	Anticalin	lipocalin	160-180	4	0 – 2 S-S	www.pieris.biz
	Fluorobody?	"superfolder" GFP	238	≥ 1	fluorophore	–
	Microbody	knottin CBD/EETI-II	36/28	(β) ¹	2/3 S-S	www.nascacell.de
	Kunitz domain	BPTI/APPI	58	1 to 2	3 S-S	www.dyax.com
2 nd ary struct.	aptamer	thioredoxin	108	1	1 S-S	–
	Affiline	γB-crystallin/ubiquitin	198	(β) ¹	–	www.scilproteins.de
	Affibody	protein A	58	2*(α) ¹	–	www.affibody.com
	DARPin	ankyrin repeat	166	3*(α/β-turn) ¹	–	www.molecularpartners.ch
oligo-mer	Tetranectin	C-type lectin domain ₃	ca. 155	3	3 S-S	www.boreanpharma.com
	Avimer	LDLR-A module _n	43	~ 4	3 + CA ²⁺	wwwext.amgen.com

(sound technology / scientific concept / patent application) (represented by consortium member)



Value of alternative scaffolds for proteome research?

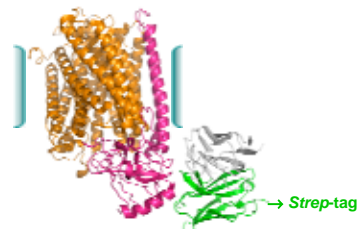


Practically:

- cost of generation (including affinity maturation) and validation
- availability of suitable "high throughput" facilities
- academic needs and capabilities vs. business requirements

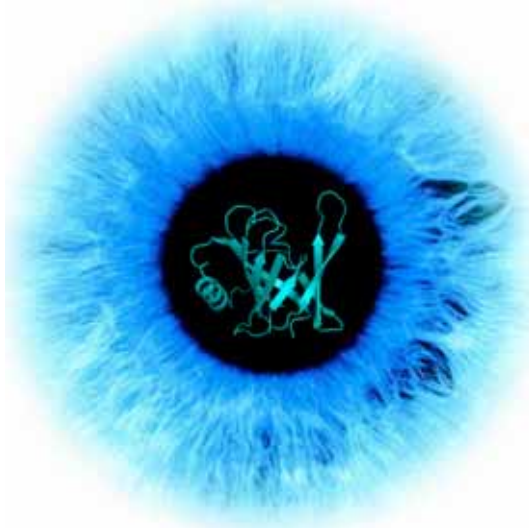
↳ Short term future in specialized and/or commercially rewarding applications?

- therapeutics [*intellectual property*]
- *in vivo* diagnostics [*plasma half life, receptor interactions*]
- highly functionalized reagents for unique applications or frequently assayed targets [*market volume*]
- crystallization tools?





Contributors



Lipocalins – Anticalins®:

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Gabriele Matschiner
Stefan Trentmann
Hendrik Gille
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and coworkers from PIERIS



Protein Crystallography:

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Daniel Breustedt
Andreas Eichinger
Dorian Schönfeld

Collaborators as mentioned